

(19) World Intellectual Property Organization
International Bureau(43) International Publication Date
13 March 2003 (13.03.2003)

PCT

(10) International Publication Number
WO 03/020756 A2

- (51) International Patent Classification⁷: C07K 14/195 (74) Agents: MARSHALL, Cameron, John et al.; Carpmals & Ransford, 43 Bloomsbury Square, London WC1A 2RA (GB).
- (21) International Application Number: PCT/IB02/03904
- (22) International Filing Date:
6 September 2002 (06.09.2002) (81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.
- (25) Filing Language: English
- (26) Publication Language: English
- (30) Priority Data:
0121591.2 6 September 2001 (06.09.2001) GB
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- (84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

[Continued on next page]

(54) Title: HYBRID AND TANDEM EXPRESSION OF NEISSERIAL PROTEINS

ΔG287-919-His	961c-741 _{MC58} -His	Orf46.1-287-His
ΔG287-Orf46.1-His	961c-983-His	Orf46.1-919-His
ΔG287-953-His	961c-Orf46.1-His	Orf46.1-741 _{MC58} -His
ΔG287-961-His	961cL-741 _{MC58}	Orf46.1-961-His
ΔG287-230-His	961cL-287	Orf46.1-961c-His
ΔG287-936-His	961c-230-His	Orf46.1-983-His
ΔG287-287-His	961c-936-His	Orf46.1-936-His
ΔG287-287 _{nz} -His		Orf46.1-230-His
ΔG287-741 _{MC58} -His		
ΔG287-741 _{ET37} -His		
		230-741 _{MC58} -His
		230-Orf46.1-His
ΔG287 _{nz} -919-His	ΔG741 _{MC58} -961c-His	230-961-His
ΔG287 _{nz} -953-His	ΔG741 _{MC58} -961-His	230-961c-His
ΔG287 _{nz} -961-His	ΔG741 _{MC58} -983-His	936-741 _{MC58} -His
ΔG287 _{nz} -287-His	ΔG741 _{MC58} -Orf46.1-His	936-Orf46.1-His
ΔG287 _{nz} -287 _{nz} -His	ΔG741 _{MC58} -741 _{MC58} -His	936-961-His
ΔG287 _{nz} -741 _{MC58} -His	ΔG741 _{MC58} -741 _{ET37} -His	936-741 _{ET37} -His
ΔG287-919-Orf46.1-His		ΔG983-741 _{MC58} -His
ΔG287-Orf46.1-919-His	919-287	ΔG983-961c-His
919-287-Orf46-His	953-287	ΔG983-961-His
Orf46.1-287-919-His	919-Orf46.1-His	ΔG983-Orf46.1-His

(57) Abstract: Two or more Neisserial proteins are joined such that they are translated as a single polypeptide chain. Hybrid proteins are represented by the formula NH₂-A-[-X-L]_n-B-COOH where X is an amino acid sequence, L is an optional linker amino acid sequence, A is an optional N-terminal amino acid sequence, B is an optional C-terminal amino acid sequence, and n is an integer greater than 1. Proteins where each of the n -X- moieties shares sequence identity to each other -X- moiety, the protein is a 'tandem protein'.

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6	2	$\Delta G287_{2996}$	(Gly) ₆	$\Delta G287_{394/98}$	-
7	2	$\Delta G287_{2996}$	(Gly) ₆	$\Delta G287_{2996}$	-
8	2	$\Delta G287_{394/98}$	(Gly) ₆	$\Delta G287_{394/98}$	-
9	2	$\Delta G287_{394/98}$	(Gly) ₆	$\Delta G287_{2996}$	-
10	2	$\Delta G741_{MC58}$	-	$741_{394/98}$	(His) ₆
11	2	$\Delta G741_{MC58}$	-	$741_{90/18311}$	(His) ₆
12	2	$\Delta G741_{MC58}$	-	741_{95N477}	(His) ₆

Proteins #1 to #5 have all been expressed in soluble form in *E.coli*. Expression levels were between 0.24 and 0.50 mg protein per litre of culture. The tandem proteins were purified and mixed with aluminium phosphate as an adjuvant. Tandem proteins #2, #4 and #5 adsorbed readily to aluminium phosphate; adsorption was less complete for tandem proteins #1 and #3.

5 Allelic variants – 741

Twenty-two polymorphic sequences of 741 were found (SEQ IDs 1 to 22). These and the MC58 sequence are aligned in Figure 1.

Allelic variants – NMB1343

Using PCR on 42 strains of meningococcus of various serogroups, the gene encoding NMB1343 protein was found in 24/42 and was absent in 18/42 strains (Table 1). The NMB1343 gene was sequenced for 10 of the NMB1343⁺ strains (Table 1, column 3). The nucleic acid sequence (and thus amino acid sequence SEQ ID 23; GenBank AAF41718) was identical in all 10 strains.

NMB1343 was also detected in two strains of *N.gonorrhoeae* (F62 and SN4). The amino acid sequence from gonococcus is SEQ ID 24. An alignment with the meningococcal sequence is:

```

15      . . . .10 . . . .20 . . . .30 . . . .40 . . . .50
      Ng   1: INNLWEISYLYRGISCCQDEQNNGQLKPKGNKAEVAIRYDGKFKYDGKAT: 50
      Nm   1: ~~~~~MGNFLYRGISCCQDEQNNGQLKPKGNKAEVAIRYDGKFKYDGKAT: 45

      . . . .60 . . . .70 . . . .80 . . . .90 . . . .100
20      Ng  51: HGPSVKNAVYAHQIETGLYDGCYISTTTDKETAKKFATSSGIENGYIYVL: 100
      Nm  46: HGPSVKNAVYAHQIETGLYDGCYISTTTDKETAKKFATSSGIENGYIYVL: 95

      . . . .110 . . . .120 . . . .130 . . . .140 . . . .150
25      Ng 101: NRDLFGQYSIFEYEVEHPENPDEKEVTIRAEDCGCIPEEVIIAKELIEIN: 150
      Nm  96: NRDLFGQYSIFEYEVEHPENPNEKEVTIRAEDCGCIPEEVIIAKELIEIN: 145

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An alignment of the corresponding nucleotide sequences is shown in Figure 2. This shows that the gonococcal sequence has a 4mer insertion in the 5' region of the NMB1343 gene which causes a frameshift and consequent loss of the 5' methionine residue.

Domain deletion – 961

961 is not present in the *N.meningitidis* serogroup A genome sequence [81], even though the surrounding regions are conserved (>90%) between serogroups A and B. References 11 and 12 disclose polymorphic forms of 961. The gene was found to be present in 91% of serogroup B strains belonging to hypervirulent lineages ET-5, ET-37 and cluster A4, but was absent in all strains of lineage 3 tested. Most of the serogroup C strains tested were positive even if not belonging to hypervirulent lineages. The same was true for the serogroup B strains with serotype 2a and 2b. For serogroup A, one strain belonging to subgroup III was positive whereas the other two strains belonging to subgroup IV-1 were negative. 961 was absent in *N.gonorrhoeae* and in commensal species *N.lactamica* and *N.cinerea*.

Figures 4 and 5 show domains in protein 961.

When the anchor region (domain 9) of protein 961 is deleted ('961cL') and expressed in *E.coli*, the protein is exported in the periplasm and secreted in the supernatant of the culture.

To investigate this further, deletion mutants in the C-terminal region of 961 were constructed (961cL-Δaro, 961cLΔcc, 961aL, 961aL-Δ1, 961aL-Δ2, 961aL-Δ3) on the basis of structural features (deletions of aromatic residues in the cases of 961cLΔaro mutant, and of coiled-coil regions for the others). These were analysed for expression and secretion into the periplasm and the supernatant of the culture. In all of these deletion mutants, the protein is produced in large amount, is present in periplasmic fraction, and is released in the supernatant of the culture.

ΔG287 – cross-strain bactericidal activity

287 was cloned for five different *N.meningitidis* serogroup B strains and was manipulated to delete the N-terminus up to the end of the poly-glycine region and to introduce a C-terminal his-tag. This gave five ΔG287 proteins. These were adjuvanted with FCA and used to raise immune sera in mice, which were then tested for bactericidal activity against all five serogroup B strains and also against serogroup A and C strains. Bactericidal titres were as follows:

Protein strain	Sera tested for bactericidal activity against strain *						
	2996	BZ232	MC58	1000	394/98	F6124	BZ133
2996	16000	128	4096	4096	1024	8000	16000
BZ232	>8000	256	2048	8000	2048	16000	8000
MC58	>8000	64	>8000	8000	2048	8000	8000
1000	>8000	64	4096	8000	1024	16000	16000
394/98	>16000	128	16000	>2048	>16000	-	-

* titres against homologous strain shown in bold

Refolding

To improve the levels of soluble protein for some hybrid proteins, alternative refolding protocols to those disclosed in reference 2 were adopted.

Inclusion bodies (IBs) were isolated as follows:

- 5 1. Homogenize cells (5g wet weight) in 25 ml 0.1 M Tris-HCl pH 7, 1mM EDTA, at 4°C using an ultraturrax (10 000 rpm)
2. Add 1.5mg lysozyme per gram cells, mix shortly with an ultraturrax, and incubate at 4°C for 30 min.
3. Use sonication or high-pressure homogenization (French press) to disrupt the cells.
- 10 4. To digest DNA, add MgCl₂ to a final concentration of 3mM and DNase to a final concentration of 10µg/ml, and incubate for 30 min at 25°C
5. Add 0.5 vol. 60 mM EDTA, 6% Triton X-100, 1,5M NaCl pH7, to the solution, and incubate for 30 min at 4°C.
6. Spin down inclusion bodies by centrifugation at 31000g (20 000 rpm) for 10 min, 4°C.
- 15 7. Resuspend pellet in 40 ml 0.1 M tris-HCl pH 7, 20mM EDTA, using an ultraturrax
8. Repeat centrifugation step 6.
9. The inclusion body pellet may be used, or stored frozen at -20°C.

Hybrid proteins were expressed in *E.coli* as follows:

Protein	Culture volume (litres)	Flask volume (litres)	Temp (°C)	Final OD ₆₀₀	Inclusion body yield (w/w)
ORF46.1-961-His	1	2	37	1.51	33.2%
ORF46.1-961c-His	1	2	37	1.6	28.3%
961c-ORF46.1His	1	2	37	1.18	23.5%
orf46.1-741 His	5	5	37	12.42	35.2

The pellets were solubilised, refolded, ultrafiltered, dialysed, and protein was then purified:

- 20 **ORF46.1-961-His** IBs were solubilised as follows: IB proteins were resuspended in 4 ml of 6M guanidine HCl, 1 mM EDTA pH 8.5 buffer, to a final protein concentration of 1 mg/ml. To refold the protein, 2 ml of solubilised protein was diluted in 400 ml of refolding buffer (0.1M Tris HCl, 1M L-arginine, 2mM EDTA pH 8.2) and incubated for 1 hour at 15°C, resulting in a protein concentration of 5µg/ml. Subsequently, another 2 ml of the solubilised protein was added and incubated for an
- 25 additional hour at the same temperature resulting in a final protein concentration of 10 µg/ml. The material was ultrafiltered using a 300 ml Amicon ultrafiltration cell (8400), applying a 3 bar pressure on an Amicon membrane with a 30 kDa cut-off (YM30) resulting in 130 ml final volume. The

ultrafiltered material was dialysed using a regenerated cellulose tubular membrane with a 12-14 kDa cutoff (Cellusep – Step bio) for 24h against 10 L of 0.1M Tris HCl pH 8,2 buffer. A second dialysis of 24h against 10 L of 300mM NaCl, 50 mM sodium phosphate pH 8,0 buffer was performed. The dialysed material was centrifuged at 22000 rpm for 45 minutes at 4°C in a Beckman centrifuge rotor JA25.5 The supernatant isolated after centrifugation was used for His-tag purification.

orf 46.1-961c-His IBs were solubilised as follows: IB proteins were resuspended in 4 ml of 6M guanidine HCl, 1 mM EDTA pH 8.5 buffer, to a final protein concentration of 1 mg/ml. To refold the protein, 2 ml of the solubilised protein was diluted in 400 ml refolding buffer (0.5M Tris HCl, 1M L-arginine, 2 mM EDTA pH 8.2) and incubated for 1 h at 15°C, resulting in a protein concentration of 5µg/ml. Subsequently another 2 ml of the solubilised protein was added and incubated for an additional hour at the same temperature resulting in a final protein concentration of 10µg/ml. The material was ultrafiltered using a 300 ml Amicon ultrafiltration cell (8400), applying a 3 bar pressure on an Amicon membrane with a 30 kDa cut-off (YM30) resulting in 150 ml final volume. The ultrafiltered material was dialysed using a regenerated cellulose tubular membrane with a 12-14 kDa cutoff (Cellusep – Step bio) for 24h against 10 L of 0.1M Tris HCl pH 8,2 buffer. A second dialysis of 24h against 10 L of 300mM NaCl, 50 mM sodium phosphate pH 8,0 buffer was performed. The dialysed material was centrifuged at 22000 rpm for 45 minutes at 4°C in a Beckman centrifuge rotor JA25.5. The supernatant isolated after centrifugation was used for His-tag purification.

961c-orf46.1-His IBs were solubilised as follows: IB proteins were resuspended in 4 ml of 6M guanidine HCl, 1 mM EDTA pH 8.5 buffer, to a final protein concentration of 1 mg/ml. To refold the protein, 2 ml of the solubilised protein was diluted in 400 ml refolding buffer (0.1M Tris HCl, 0.5 M L-arginine, 2 mM EDTA pH 8.2) and incubated for 1 h at 15°C, resulting in a protein concentration of 5 µg/ml. Subsequently another 2 ml of the solubilized protein was added and incubated for an additional hour at the same temperature resulting in a final protein concentration of 10 µg/ml. The material was ultrafiltered using a 300 ml Amicon ultrafiltration cell (8400), applying a 3 bar pressure on an Amicon membrane with a 30 kDa cut-off (YM30) resulting in 150 ml final volume. The ultrafiltered material was dialysed using a regenerated cellulose tubular membrane with a 12-14 kDa cutoff (Cellusep – Step bio) for 24h against 10 L of 0.1M Tris HCl pH 8,2 buffer. A second dialysis of 24h against 10 L of 300mM NaCl, 50 mM sodium phosphate pH 8,0 buffer was performed. The dialysed material was centrifuged at 22000 rpm for 45 minutes at 4°C in a Beckman centrifuge rotor JA25.5. The supernatant isolated after centrifugation was used for His-tag purification.

orf46.1-741-His IBs were solubilised as follows: IB proteins were resuspended in 4 ml of 6M guanidine HCl, 1 mM EDTA pH 8.5 buffer, to a final protein concentration of 10 mg/ml. To refold, 2 ml of the solubilised protein was diluted in 400 ml of the refolding buffer (0.5M Tris HCl, 0.7 M L-arginine, 2 mM EDTA pH 7.2) and incubated for 1 h at 15°C, resulting in a protein concentration of

50µg/ml. Subsequently another 2 ml of the solubilised protein was added and incubated for an additional hour at the same temperature resulting in a final protein concentration of 100µg/ml. The material was ultrafiltered using a 300 ml Amicon ultrafiltration cell (8400), applying a 3 bar pressure on an Amicon membrane with a 30 kDa cut-off (YM30) resulting in 120 ml final volume. The ultrafiltered material was dialysed using a regenerated cellulose tubular membrane with a 12-14 kDa cutoff (Cellusep – Step bio) for 24h against 10 L of 0.1M Tris HCl pH 8,2 buffer. A second dialysis of 24h against 10 L of 300mM NaCl, 50 mM sodium phosphate pH 8,0 buffer was performed. The dialysed material was centrifuged at 22000 rpm for 45 minutes at 4°C in a Beckman centrifuge rotor JA25.5. The supernatant isolated after centrifugation was used for His-tag purification.

- 10 Compared with proteins purified as described in ref. 2, bactericidal assay titres were as follows:

Protein	Reference 2		Refolded		
	CFA	Aluminium hydroxide	Aluminium hydroxide	MF59	Aluminium phosphate
ORF46.1-961-His	8192	8192	32768	-	-
ORF46.1-961c-His	8192	128	<64	8192	-
961c-ORF46.1His	32768	1024	16384	-	-
orf46.1-741 His	<4	16	<4	256	-

Similar procedures were used for ORF46.1 to purify the protein from IBs when expressed with no His-tag ('ORF46.1K'):

Protein	Culture volume (litres)	Flask volume (litres)	Temp (°C)	Final OD ₆₀₀	Inclusion body yield (w/w)
orf46.1K	5	5	37	13.7	29.4

- IB proteins were resuspended in 4 ml of 6M guanidine HCl, 1 mM EDTA pH 8.5 buffer, to a final protein concentration of 10 mg/ml. To refold, 2 ml of the solubilised protein was diluted in 400 ml of the refolding buffer (0.5M Tris HCl, 0.7 M L-arginine, 2 mM EDTA pH 7.2) and incubated for 1 hours at 15°C, resulting in a protein concentration of 50µg/ml. Subsequently another 2 ml of the solubilised protein was added and incubated for an additional hour at the same temperature resulting in a final protein concentration of 100µg/ml. The material was ultrafiltered using a 300ml Amicon ultrafiltration cell (8400), applying a 3 bar pressure on an Amicon membrane with a 30kDa cut-off (YM30) resulting in 120 ml final volume. The ultrafiltered material was dialysed using a regenerated cellulose tubular membrane with a 12-14 kDa cutoff (Cellusep – Step bio) for 12h against 10 L of 50mM sodium phosphate, 2mM EDTA, pH 7,2 buffer. A second dialysis of 24h against 10 L of the same buffer was performed. The dialysed material was centrifuged at 22000 rpm for 45 minutes at 4°C in a Beckman centrifuge rotor JA25.5. The supernatant isolated after centrifugation was used for cationic exchange chromatography. The purification was done on a AKTA explorer chromatography

system (Amersham-Pharmacia Biotech) using a 5 ml HiTrap SP sepharose HP column (Amersham-Pharmacia Biotech). The flow rate applied was of 1.5 ml per minute. The column was washed with 35 ml of 50mM sodium phosphate buffer pH 7.2. A linear gradient (0-1 M NaCl) was performed using a 50mM sodium phosphate buffer pH 7.2. The protein eluted in two peaks at 92 mM and 380mM NaCl. The fractions constituting each peak were pooled and respectively named pool 1 and pool 2.

Compared with proteins purified as described in ref. 2, bactericidal assay titres when adjuvanted with aluminium hydroxide were improved from <4 to 1024. The titre using aluminium phosphate adjuvant with the refolded protein was 2048. ELISA titres were as follows:

Protein	Aluminium adjuvant	Elisa (M7)	SBA (2996)
Orf46.1k (pool 1)	Hydroxide 3.3mg/ml	1212	512
	Phosphate 0.6 mg/ml	154	1024
Orf46.1k (pool 2)	Hydroxide 3.3mg/ml	1085	1024
	Phosphate 0.6 mg/ml	250	1024

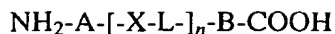
- 10 It will be understood that the invention has been described by way of example only and modifications may be made whilst remaining within the scope and spirit of the invention.

TABLE 1

Strain	1343	Sequence	Strain classification
72/00	+		ET5 B:15:P1.7,13,13a
30/00	+		ET5 B:15:P1.7,16
39/99	+		ET5 C:15:P1.7,16
95330	+		ET5 B:4:P1.15
M4102	+		ET5 nd
MC58(21)	+	+	ET5 B:15:P1.7,16b
BZ169(7)	+	+	ET5 B:NT:P1.16
BZ83(19)	+		ET5 B:15:-.-
CU385	+	+	ET5 B:4:P1.15
220173I	+		ET5 NG:4:P1.15
64/96	+	+	ET5 NG:15:P1.7,16 (carrier)
220173I	+		ET5 B:4:P1.15 (carrier)
ISS1071	+		nd B:15:P1.7,16 (ET5?)
BZ198(2)	+	+	lin.3 B:8:P1.1
980-2543	+	+	lin.3 B:NT:P1.4
16060	+	+	other B:4:P1.14 (carrier)
394-98	+		nd B:4:P1.4 (lin 3?)
ISS1106	+		nd B:4:P1.4 (lin.3?)
BZ133(10)	+	+	sub I B:NT:-.-
S3446	+	+	nd B:14:P1.23,14
ISS1001	+	+	nd B:14:P1.13
241175I	+		other NG:21:P1.16 (carrier)
171274I	+		other NG:15:- (carrier)
66/96	+		other B:17:P1.15 (carrier)
961-5945	-		A4
96217	-		A4
312294	-		A4
90/18311(24)	-		ET37
93/4286(25)	-		ET37
M986	-		ET37
1000(5)	-		other
NGE28(13)	-		other carrier
NGH38(14)	-		other carrier
BZ232(18)	-		other
F6124(23)	-		sub III A:-.-
C11	-		C:-
NMB	-		nd
8047	-		nd
ISS759	-		nd C:2b:P1.2
ISS1113	-		nd C:2:P1.5
65/96	-		nd 4:P1.14
2996(96)	-		nd B:2b:P1.5,2

CLAIMS

1. A hybrid protein having formula:



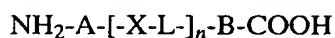
wherein L is an optional linker amino acid sequence, A is an optional N-terminal amino acid sequence, B is an optional C-terminal amino acid sequence, and n is an integer greater than 1, and X is either:

(a) an orf1, orf4, orf25, orf40, orf46.1, orf83, NMB1343, 230, 233, 287, 292, 594, 687, 736, 741, 907, 919, 936, 953, 961 or 983 amino acid sequence;

(b) an amino acid sequence having sequence identity to an amino acid sequence from (a); or

(c) an amino acid sequence comprising a fragment of an amino acid sequence from (a).

2. A hybrid protein having formula:



wherein X is an amino acid sequence, L is an optional linker amino acid sequence, A is an optional N-terminal amino acid sequence, B is an optional C-terminal amino acid sequence, n is an integer greater than 1, and wherein a first X moiety ($-\text{X}_a-$) has one of the following amino acid sequences:

(d) the 446 even SEQ IDs (i.e. 2, 4, 6, ..., 890, 892) disclosed in reference 3.

(e) the 45 even SEQ IDs (i.e. 2, 4, 6, ..., 88, 90) disclosed in reference 4;

(f) the 1674 even SEQ IDs 2-3020, even SEQ IDs 3040-3114, and all SEQ IDs 3115-3241, disclosed in reference 5;

(g) the 2160 amino acid sequences NMB0001 to NMB2160 from reference 7; or

(h) an amino acid sequence disclosed in reference 1 or reference 2,

and a second $-\text{X}-$ moiety ($-\text{X}_b-$), wherein $-\text{X}_b-$ has sequence identity to $-\text{X}_a-$ and/or $-\text{X}_b-$ comprises a fragment of $-\text{X}_a-$.

3. The hybrid protein of claim 1 or claim 2, wherein $n=2$.

4. The hybrid protein of claim 2, wherein $-\text{X}_a-$ is an orf46.1, 230, 287, 741, 919, 936, 953, 961 or 983 amino acid sequence.

5. The hybrid protein of claim 2, wherein $\text{X}_1, \dots, \text{X}_n$ all have sequence identity to each other.

6. The hybrid protein of any preceding claim, wherein $n=2$, and wherein the $-\text{X}-$ moieties are: ΔG287 and 230; ΔG287 and 936; ΔG287 and 741; 961c and 287; 961c and 230; 961c and 936;

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961cL and 287; 961cL and 230; 961cL and 936; ORF46.1 and 936; ORF46.1 and 230; 230 and 961; 230 and 741; 936 and 961; 936 and 741; Δ G741 and 741; or Δ G287 and 287.

7. The hybrid protein of any preceding claim, wherein L has 20 or fewer amino acids.
8. The hybrid protein of any preceding claim, wherein L is a poly-glycine linker.
- 5 9. The hybrid protein of any preceding claim, wherein A has 40 or fewer amino acids.
10. The hybrid protein of any preceding claim, wherein B has 40 or fewer amino acids.
11. The hybrid protein of any preceding claim, wherein the -X- moieties have an amino acid sequence found in *N.meningitidis* serogroup B.
12. The hybrid protein of any preceding claim, wherein at least one -X- moiety is a 961 amino acid sequence in which one or more domains has been deleted.
- 10 13. A protein comprising an amino acid sequence selected from the group consisting of SEQ IDs 1 to 38.
14. Nucleic acid encoding a protein of any preceding claim.
15. A composition comprising protein or nucleic acid according to any preceding claim.
- 15 16. A composition comprising two or more of the following proteins:
 - (1) 287
 - (2) 741
 - (3) ORF46.1
 - (4) 961
 - 20 (5) $\text{NH}_2\text{-A-}[-\text{X-L}]_n\text{-B-COOH}$, wherein $n=2$, $\text{X}_1=287$, $\text{X}_2=953$
 - (6) $\text{NH}_2\text{-A-}[-\text{X-L}]_n\text{-B-COOH}$, wherein $n=2$, $\text{X}_1=287$, $\text{X}_2=919$
 - (7) $\text{NH}_2\text{-A-}[-\text{X-L}]_n\text{-B-COOH}$, wherein $n=2$, $\text{X}_1=287$, $\text{X}_2=961$
 - (8) $\text{NH}_2\text{-A-}[-\text{X-L}]_n\text{-B-COOH}$, wherein $n=2$, $\text{X}_1=287$, $\text{X}_2=741$
 - (9) $\text{NH}_2\text{-A-}[-\text{X-L}]_n\text{-B-COOH}$, wherein $n=2$, $\text{X}_1=936$, $\text{X}_2=741$
- 25 17. The composition of claim 16, comprising proteins (4), (5) and (9).
18. The composition of claim 17, wherein protein (4) comprises SEQ ID 31, protein (5) comprises SEQ ID 28 or SEQ ID 29, and protein (9) comprises SEQ ID 30.
19. The composition of any one of claims 15 to 18, further comprising:
 - a protein antigen from *N.meningitidis*;
 - 30 – an outer-membrane vesicle (OMV) preparation from *N.meningitidis*;
 - a saccharide antigen from *N.meningitidis*;
 - a saccharide antigen from *Streptococcus pneumoniae*;

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- an antigen from hepatitis A, B or C virus;
 - an antigen from *Bordetella pertussis*;
 - a diphtheria antigen;
 - a tetanus antigen;
 - 5 – a protein antigen from *Helicobacter pylori*;
 - a saccharide antigen from *Haemophilus influenzae*;
 - an antigen from *N.gonorrhoeae*;
 - an antigen from *Chlamydia pneumoniae*;
 - an antigen from *Chlamydia trachomatis*;
 - 10 – an antigen from *Porphyromonas gingivalis*;
 - polio antigen(s);
 - rabies antigen(s);
 - measles, mumps and/or rubella antigens;
 - influenza antigen(s);
 - 15 – an antigen from *Moraxella catarrhalis*;
 - an antigen from *Streptococcus agalactiae*;
 - an antigen from *Streptococcus pyogenes*; and/or
 - an antigen from *Staphylococcus aureus*.
20. The composition of any one of claims 15 to 19, further comprising a pharmaceutically acceptable
20 carrier.
21. The composition of claim 20 for use as a medicament.
22. A method of treating a patient, comprising administering to the patient a therapeutically effective amount of the composition of claim 20.

SEQUENCE LISTING***SEQ ID 1 – 741 from strain 1000***

MTRSKPVNRTAFCCSLTAALILTACSSGGGGVAADIGAGLADALTTPLDHKDKSLQSLTLDQSVRKNEK
 LKLAAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQTITLASGEFQIYKQNHSAVVALQIEKIN
 5 NPDKIDSLINQRSFLVSGLGGEHTAFNQLPDGKAEYHGKAFSSDDPNGLRLHYSIDFTKKQGYGRIEHLKT
 PEQNVELASAEKKADEKSHAVILGDTRYGGEEKGTYHLALFGDRAQEIAGSATVKIREKVHEIGIAGKQ

SEQ ID 2 – 741 from strain 2201731 (premature stop codon, though reliable sequence)

MTRSKPVNRTAFCCSLTTALILTACSSGGGGVAADIGAGLADALTAPLDHKDKGLQSLTLDQSVRKNEK
 LKLAAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQVYKQSHSALTAFQTEQIQ
 10 DSEHSGKMVAKRQFRIGDIAGEHTSFDKLPPEGGRATYRGTAFGSDDAGGKLTYTIDFAAQQNGKIEHLK
 SPELNVDLAAADIKPDGKRHAVISGSVLYNQAEGSYSLGIFGGKA

SEQ ID 3 – 741 from strain 90/18311 (incomplete)

GLADALTAPLDHKDKSLQSLTLDQSVRKNEKLKLAAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEV
 DGQLITLESGEFQIYKQDHSAVVALQIEKINNPDKIDSLINQRSFLVSGLGGEHTAFNQLPSGKAHYHGK
 15 AFSSDDPNGLRLHYSIDFTKKQGYGRIEHLKTPEQNVELASAEKKADEKSHAVILGDTRYGGEEKGTYHLA
 LFGDRAQEIAGSATVKIREKVHET

SEQ ID 4 – 741 from strain L93/4286 (incomplete)

VAADIGAGLADALTAPLDHKDKGLQSLMLDQSVRKNEKLKLAAQGAEKTYGNGDSLNTGKLKNDKVSFRD
 FIRQIEVDGQTITLASGEFQIYKQNHSAVVALQIEKINNPDKIDSLINQRSFLVSGLGGEHTAFNQLPDG
 20 KAEYHGKAFSSDDPNGLRLHYSIDFTKKQGYGRIEHLKTPEQNVELASAEKKADEKSHAVILGDTRYGGE
 KGTYHLALFGDRAQEIAGSATVKIREKVHEIGIAGKQ

SEQ ID 5 – 741 from strain 2996

MTRSKPVNRTAFCCSLTAALILTACSSGGGGVAADIGAGLADALTAPLDHKDKSLQSLTLDQSVRKNEK
 LKLAAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQIYKQDHSAVVALQIEKIN
 25 NPDKIDSLINQRSFLVSGLGGEHTAFNQLPDGKAEYHGKAFSSDDAGGKLTYTIDFAAQQGHGKIEHLKT
 PEQNVELAAAEKKADEKSHAVILGDTRYGSEKGTYHLALFGDRAQEIAGSATVKIGEKVHEIGIAGKQ

SEQ ID 6 – 741 from strain 30/00

KDKGLQSLTLDQSVRKNEKLKLAAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEF
 QVYKQSHSALTAFQTEQIQDSEHSGKMVAKRQFRIGDIAGEHTSFDKLPPEGGRATYRGTAFGSDDAGGKL
 30 TYTIDFAAQQNGKIEHLKSPELNVDLAAADIKPDGKRHAVISGSVLYNQAEGSYSLGIFGGKAQEVAG
 SAEVKTVNGIRHIGLAAKQ

SEQ ID 7 – 741 from strain 312294

MTRSKPVNRTAFCCSLTAALILTACSSGGGGVAADIGAGLADALTAPLDHKDKSLQSLTLDQSVRKNEK
 LKLAAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQIYKQDHSAVVALQIEKIN
 35 NPDKIDSLINQRSFLVSGLGGEHTAFNQLPDGKAEYHGKAFSSDDAGGKLTYTIDFAAQQGHGKIEHLKT
 PEQNVELAAAEKKADEKSHAVILGDTRYGSEKGTYHLALFGDRAQEIAGSATVKIGEKVHEIGIAGKQ

SEQ ID 8 – 741 from strain 39/99 (incomplete)

DKGLQSLTLDQSVRKNEKLKLAAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQ
 VYKQSHSALTAFQTEQIQDSEHSGKMVAKRQFRIGDIAGEHTSFDKLPPEGGRATYRGTAFGSDDAGGKLT
 40 YTIDFAAQQNGKIEHLKSPELNVDLAAADIKPDGKRHAVISGSVLYNQAEGSYSLGIFGGKAQEVAGS
 AEVKTVNGIRHIGLAAKQ

SEQ ID 9 – 741 from strain 5/99

MTRSKPVNRTAFCCSLTAALILTACSSGGGGVAADIGAGLADALTAPLDHKDKGLQSLTLDQSVRKNEK
 LKLAAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQIYKQDHSAVVALQIEKIN
 45 NPDKIDSLINQRSFLVSGLGGEHTAFNQLPSGKAHYHGKAFSSDDPNGLRLHYSIDFTKKQGYGRIEHLKT
 PEQNVELASAEKKADEKSHAVILGDTRYGGEEKGTYHLALFGDRAQEIAGSATVKIREKVHEIGIAGKQ

SEQ ID 10 – 741 from strain 67/00

MTRSKPVNRTAFCCFSLTAALILTACSSGGGGVAADIGAGLADALTAPLDHKDKSLQSLTLDQSVRKNEK
LKLAQAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQVYKQSHSALTALQTEQEQ
DPEHSGKMAKRRFKIGDIAGEHTSFDKLPKDVMTYRGTAFGSDDAGGKLTYTIDFAAQQGHGKIEHLK
5 SPELNVELATAYIKPDEKHHAVISGSVLYNQDEKGSYSLGIFGGQAQEVAGSAEVTANGIHHIGLAAQ

SEQ ID 11 – 741 from strain BZ169

LQSLTLDQSVRKNEKLKLAQAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQVYK
QSHSALTAFQTEQIQDSEHSGKMAKRRQFRIGDIAGEHTSFDKLPPEGGRATYRGTAFGSDDAGGKLTITI
DFAAQQNGKIEHLKSPELNVDLAAADIKPDGKRHAVISGSVLYNQAEKGSYSLGIFGGKAQEVAGSAEV
10 KTVNGIRHIGLAAQ

SEQ ID 12 – 741 from strain 72/00

LQSLTLDQSVRKNEKLKLAQAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQVYK
QSHSALTAFQTEQIQDSEHSGKMAKRRQFRIGDIAGEHTSFDKLPPEGGRATYRGTAFGSDDAGGKLTITI
DFAAQQNGKIEHLKSPELNVDLAAADIKPDGKRHAVISGSVLYNQAEKGSYSLGIFGGKAQEVAGSAEV
15 KTVNGIRHIGLAAQ

SEQ ID 13 – 741 from strain 93/114

MTRSKPVNRTAFCCFSLTAALILTACSSGGGGVAADIGAGLADALTAPLDHKDKSLQSLTLDQSVRKNEK
LKLAQAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQVYKQSHSALTALQTEQEQ
DPEHSGKMAKRRFKIGDIAGEHTSFDKLPKDVMTYRGTAFGSDDAGGKLTYTIDFAAQQGHGKIEHLK
20 SPELNVELATAYIKPDEKHHAVISGSVLYNQDEKGSYSLGIFGGQAQEVAGSAEVTANGIHHIGLAAQ

SEQ ID 14 – 741 from strain 95N477

MTRSKPVNRTAFCCSLTAALILTACSSGGGGVAADIGAGLADALTAPLDHKDKSLQSLTLDQSVRKNEK
LKLAQAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQIYKQDHSAVVALQIEKIN
NPDKIDSLINQRSFLVSGLGGEHTAFNQLPSGKAIEYHGKAFSSDDPNRGLHYSIDFTKKQGYGRIEHLKT
25 PEQNVELASAELKADEKSHAVILGDTRYGEEKGTIYHLALFGDRAQEIAGSATVKIREKVHEIGIAGKQ

SEQ ID 15 – 741 from strain 96217

MTRSKPVNRTAFCCSLTAALILTACSSGGGGVAADIGAGLADALTAPLDHKDKSLQSLTLDQSVRKNEK
LKLAQAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQIYKQDHSAVVALQIEKIN
NPDKIDSLINQRSFLVSGLGGEHTAFNQLPDGKAIEYHGKAFSSDDAGGKLTYTIDFAAQQGHGKIEHLKT
30 PEQNVELAAAELKADEKSHAVILGDTRYGSEEEKGTIYHLALFGDRAQEIAGSATVKIGIEKVHEIGIAGKQ

SEQ ID 16 – 741 from strain BZ133

MTRSKPVNRTAFCCSLTAALILTACSSGGGGVAADIGAGLADALTAPLDHKDKSLQSLTLDQSVRKNEK
LKLAQAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQVYKQSHSALTALQTEQVQ
DSEHSGKMAKRRQFRIGDIAGEHTSFDKLPPEGGRATYRGTAFGSDDASGKLTYTIDFAAQQGHGKIEHLK
35 SPELNVDLAAADIKPDKRRHAVISGSVLYNQAEKGSYSLGIFGGQAQEVAGSAEVTANGIRHIGLAAQ

SEQ ID 17 – 741 from strain BZ232

MTRSKPVNRTAFCCSLTAALILTACSSGGGGVAADIGAGLADALTAPLDHKDKSLQSLTLDQSVRKNEK
LKLAQAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQTITLASGEFQIYKQNHSAVVALQIEKIN
NPDKIDSLINQRSFLVSGLGGEHTAFNQLPDGKAIEYHGKAFSSDDPNRGLHYSIDFTKKQGYGRIEHLKT
40 PEQNVELASAELKADEKSHAVILGDTRYGEEKGTIYHLALFGDRAQEIAGSATVKIREKVHEIGIAGKQ

SEQ ID 18 – 741 from strain C11

MTRSKPVNRTAFCCSLTAALILTACSSGGGGVAADIGAGLADALTAPLDHKDKSLQSLTLDQSVRKNEK
LKLAQAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQIYKQDHSAVVALQIEKIN
NPDKIDSLINQRSFLVSGLGGEHTAFNQLPSGKAIEYHGKAFSSDDPNRGLHYSIDFTKKQGYGRIEHLKT
45 PEQNVELASAELKADEKSHAVILGDTRYGEEKGTIYHLALFGDRAQEIAGSATVKIREKVHEIGIAGKQ

SEQ ID 19 – 741 from strain M1090

MTRSKPVNRTAFCCSLTAALILTACSSGGGGVAADIGAGLADALTAPLDHKDKSLQSLTLDQSVRKNEK
LKLAQAQGAEKTYGNGDSLNTGKLKNDKVSFRDFIRQIEVDGQLITLESGEFQIYKQDHSAVVALQIEKIN

NPDKIDSLINQRSFLVSGLGGEHTAFNQLPSGKAEYHGKAFSSDDAGGKLTYYTIDFAAKQGHGKIEHLKT
PEQNVELASAELKADEKSHAVILGDTRYGGEKGTYHLALFGDRAQEIAGSATVKIREKVHEIGIAGKQ

SEQ ID 20 – 741 from strain M1096

5 MTRSKPVNRTAFCCSLTAALILTACSSGGGGVAADIGAGLADALTTPLDHKDKSLQSLTLDQSVRKNEK
LKLAQAQGAEKTYGNGDSLNTGKLNKDKVSRFDFIRQIEVDGQTITLASGEFQIYKQNHSAVVALQIEKIN
NPDKIDSLINQRSFLVSGLGGEHTAFNQLPDGKAEYHGKAFSSDDPNRLHYSIDFTKKQGYGRIEHLKT
PEQNVELASAELKADEKSHAVILGDTRYGGEKGTYHLALFGDRAQEIAGSATVKIREKVHEIGIAGKQ

SEQ ID 21 – 741 from strain M198/172

10 MTRSKPVNRTAFCCSLTAALILTACSSGGGGVAADIGAGLADALTAPLDHKDKSLQSLTLDQSVRKNEK
LKLAQAQGAEKTYGNGDSLNTGKLNKDKVSRFDFIRQIEVDGQLITLESGEFQVYKQSHSALTALQTEQVQ
DSEHSGKMAKVRQFRIGDIAGEHTSFDKLEGGGRATYRGTAFGSDDASGKLTYYTIDFAAKQGHGKIEHLK
SPELNVDLAASDIKPKKRHAVISGSVLYNQAEGSYSLGIFGGQAQEVAGSAEVETANGIRHIGLAQKQ

SEQ ID 22 – 741 from strain NGH38

15 MTRSKPVNRTAFCCSLTAALILTACSSGGGGVAADIGAGLADALTAPLDHKDKSLQSLTLDQSVRKNEK
LKLAQAQGAEKTYGNGDSLNTGKLNKDKVSRFDFIRQIEVDGQTITLASGEFQIYKQNHSAVVALQIEKIN
NPDKIDSLINQRSFLVSGLGGEHTAFNQLPDGKAEYHGKAFSSDDPNRLHYSIDFTKKQGYGRIEHLKT
PEQNVELASAELKADEKSHAVILGDTRYGGEKGTYHLALFGDRAQEIAGSATVKIREKVHEIGIAGKQ

SEQ ID 23 – NMB1343 from ten meningococcal strains

20 MGNFLYRGISCCQDEQNNGQLKPKGNKAEVAIRYDGFYDYGKATHGPSVKNAVYAHQIETGLYDGCYIS
TTTDKEIAKKFATSSGIENGYIYVLNRDLFGQYSIFEYEVEHPENPNEKEVTIRAEDCGCIPEEVIIAKE
LIEIN

SEQ ID 24 – NMB1343 from gonococcus

25 INNWLWEISYLYRGISCCQDEQNNGQLKPKGNKAEVAIRYDGFYDYGKATHGPSVKNAVYAHQIETDLVD
GCYISTTTTDKEIAKKFATSSGIENGYIYVLNRDLFGQYSIFEYEVEHPENPDEKEVTIRAEDCGCIPEEV
IIAKELIEIN

SEQ ID 25 – NMB1343 nucleic acid sequence (gonococcus)

30 TCCGCCGCATTACCTTATAAAATAAAACATCCCTCTCAAGCAGTCTGATAATGTTTGGATTGCTTGAGAT
TGATGAGTGATGGTGTTAAATTCAAACCTTTAAATTAATAAATTATGGGAAATTTCTTATTTATATAGAGG
CATAGTTGCCAACAAAGATGAGCAAAATAATGGACAGTTAAACCTAAAGGTAATAAAGCTGAAGTTGCA
ATTCGTTATGATGGTAAGTTTAAATATGATGGTAAGCTACACATGGTCCAAGTGTGAAGAATGCAGTTT
ACGCCCATCAAATTGAAACAGATCTATATGACGGATGTTATATATCTACGACAACAGACAAGGAAATTGC
CAAGAAATTTGCAACAAGCTCCGGCATCGAAAATGGCTATATATATGTTTTAAATAGAGATTTGTTTGGT
CAATATTCTATTTTGAATATGAGGTTGAACATCCAGAAAACCCAGATGAGAAGGAAGTAACAATCAGAG
CTGAAGATTGTGGCTGTATTCTGAAGAAGTGATTATTGCTAAAGAGTTGATAGAAATTAACATAAGTTGA
35 AAGGTCAATATAATGGCTTTAGTTGAATTGAAAGTGCCCGACATTGGCGGACACGAAAATGTAGATATTA
TCGC

SEQ ID 26 – NMB1343 nucleic acid sequence (meningococcus)

40 TCCGCCGCATTACCTTATAAAATAAAACATCCCTCTCAAGCAGTCTGATAATGTTTGGATTGCTTGAGAT
TGATGAGTAATGGTGTTAAATTCAACCTTTAAATTAATAAATTATGGGAAATTTCTTATATAGAGGCATT
AGTTGCCAACAAAGATGAGCAAAATAATGGACAGTTAAACCTAAAGGTAATAAAGCTGAAGTTGCAATTC
GTTATGATGGTAAGTTTAAATATGATGGTAAGCTACACATGGTCCAAGTGTGAAGAATGCAGTTTACGC
CCATCAAATTGAAACAGGTCTATATGACGGATGTTATATATCTACGACAACAGACAAGGAAATTGCCAAG
AAATTTGCAACAAGTTCCGGCATCGAAAATGGCTATATATATGTTTTAAATAGGGATTTGTTTGGTCAAT
ATTCTATTTTGAATATGAGGTTGAACATCCAGAAAACCCAAATGAGAAGGAAGTAACAATCAGAGCTGA
45 AGATTGTGGCTGTATTCTGAAGAAGTGATTATTGCTAAAGAGTTGATAGAAATTAACATAAGTTGAAAGG
TCAATATAATGGCTTTAGTTGAATTGAAAGTGCCCGACATTGGCGGACACGAAAATGTAGATATTATCGC

SEQ ID 27 – linker

GSGGGG

SEQ ID 28 – preferred ΔG287-953 hybrid

MASPDVKSADTLSKPAAPVVAEKETEVEKEDAPQAGSQGGGAPSTQGSQDMAAVSAENTGNNGAATTDKPK
NEDEGPQNDMPQNSAESANQTGNNQPADSSDSAPASNPAPANGGSNFRVLDLANGVLIDGPSQNTLTHC
KGDSCNGDNLDEEAPSKSEFENLNESEKIEKYKDKGSDKFTNLVATAVQANGTNKYVIIYKDKSASSS
5 SARFRRSARSRRSLPAEMPLIPVNQADTLIVDGEAVSLTGHSGNIFAPEGNRYRLTYGAEKLPGGSYALR
VQGEPAKGEMLAGTAVYNGEVLHFHTENGRPYPTRGRFAAKVDFGSKSVDCIIDSDDLHMGQTQKFKAAI
DGNGFKGTWTENGDDVSGRFYGPAGEEVAGKYSYRPTDAEKGFGVFAGKKEQDGSGGGGATYKVDEYH
ANARFAIDHFNTSTNVGGFYGLTGSVEFDQAKRDGKIDITIPVANLQSGSQHFTDHLKSADIFDAAQYPD
IRFVSTKFNFGKLVSDGNLTMHGKTAPVKLKAKEFNQYQSPMAKTEVCGGDFSTTIDRTKWGVVDYLV
10 NVGMTKSVRIDIQIEAAKQ

SEQ ID 29 – ΔG287_{NZ}-953 hybrid

MASPDVKSADTLSKPAAPVVSEKETEAKEDAPQAGSQGGGAPSAQGGQDMAAVSEENTGNNGAAATTDKPK
NEDEGAQNDMPQNAADTDSLTPNHTPASNMPAGNMENQAPDAGESEQPANQPDMANTADGMQGGDPSAGG
ENAGNTAAQGTNQAENNQTAGSQNPASSTNPSATNSGGDFGRTNVGNVVIDGPSQNTLTHCKGDSCSG
15 NNFLDEEVQLKSEFEKLSADKISNYKKDGNKNDKDFVGLVADSVQMKGINQYIIIFYKPKPTSFARFR
RSARSRRSLPAEMPLIPVNQADTLIVDGEAVSLTGHSGNIFAPEGNRYRLTYGAEKLPGGSYALRVQGE
SKGEMLAGTAVYNGEVLHFHTENGRPSPSRGRFAAKVDFGSKSVDCIIDSDDLHMGQTQKFKAAIDGNGF
KGTWTENGDDVSGRFYGPAGEEVAGKYSYRPTDAEKGFGVFAGKKEQDGSGGGGATYKVDEYHANARF
AIDHFNTSTNVGGFYGLTGSVEFDQAKRDGKIDITIPVANLQSGSQHFTDHLKSADIFDAAQYPDIFV
20 TKFNFGKLVSDGNLTMHGKTAPVKLKAKEFNQYQSPMAKTEVCGGDFSTTIDRTKWGVVDYLVNVMGT
KSVRIDIQIEAAKQ

SEQ ID 30 – 936-ΔG741 hybrid

MKPKPHTVRTLIAAIFSLALSGCVSAVIGSAAVGAKSAVDRRRTTGAQTDDNVMALRIETTARSYLRQNNQ
TKGYTPQISVVGYNRHLALLGQVATEGEKQFVGQIARSEQAAEGVYNYITVASLPRTAGDIAGDTWNTSK
25 VRATLLGISPATQARVKIVTYGNVTVMGILTPEEQAQITQKVSTTVGVQKVITLYQNYVQRGSGGGGVA
ADIGAGLADALTAPLDHKDKGLQSLTLDQSVRKNEKLKLAQAQGAEKTYGNGDSLNTGKLKNDKVSFRDFI
RQIEVDGQLITLESGEFQVYKQSHSALTAFQTEQIQDSEHSGMKVAKRQFRIGDIAGEHTSFDKLPEGGR
ATYRGTAFGSDDAGGKLTYTIDFAAQGNGKIEHLKSPELNVDLAAADIKPDGKRHAVISGSVLYNQAEK
GSYSLGIFGGKAQEVAGSAEVKTVNGIRHIGLAAKQ

SEQ ID 31 – 961c

MATNDDDVKKAATVAIAAAYNNGQEINGFKAGETIYDIDEDGTITKKDATAADVEADDFKGLGLKKVVTN
LTKTVNENKQNVDAKVAAESEIEKLTTKLADTDAALADTDAALDATTNALNKLGENITTFAEETKTNI
KIDEKLEAVADTVDKHAEAFNDIADSLDETNTKADEAVKTANEAKQTAEETKQNVDAKVAAETAAGKAE
AAAGTANTAADKAEAVAAKVTDIKADIATNKDNIAKKANSADVTTREESDSKFVRIDGLNATTEKLDTRL
35 ASAEKSIADHDTRLNGLDKTVSDLRKETRQGLAEQAALSGLFQPYNVG